

Comparison of the effects of pulsed and continuous wave light on axonal regeneration in a rat model of spinal cord injury

Xingjia Wu · Helina Moges · Luis DeTaboada · Juanita Anders

Received: 8 March 2011 / Accepted: 3 August 2011
© Springer-Verlag London Ltd 2011

Background

Light therapy (LT) has been investigated as a viable treatment for injuries and diseases of the central nervous system in both animal models and clinical trials. Based on *in vivo* studies, LT has beneficial effects on the treatment of spinal cord injury (SCI) [1–3], traumatic brain injury [4], stroke [5–9] and neurodegenerative diseases [10, 11]. Recent clinical trials on stroke have demonstrated that light is safe for the treatment of ischemic stroke in humans [12, 13].

The biomodulatory effects of LT have been investigated to a much greater extent with continuous wave (CW) mode than in pulsed mode. However, there is no clear consensus that one mode is superior to the other. A recent review of the published literature that compared the effects of light in CW or pulsed mode reported that some studies have shown pulsed light to be more effective than CW light, while other studies have shown no effect or a worsening effect with pulsed light compared to no treatment [14]. The studies in which pulsed light was more effective than CW light include LT on wound healing, pain attenuation, bone stimulation and ischemic stroke in animal models and human studies [14]. Lapchak et al. compared CW light with two different frequencies of pulsed light and found that transcranial treatment with both pulsing regimes significantly improved

behavioral deficits compared to the CW light [6]. Why pulsed light in some cases improves the outcome of LT is currently unclear, and further investigation is needed.

Our previous studies have shown that LT promotes axonal regeneration and functional recovery in both hemisection and contusion models of SCI in rats with CW laser light [2, 3]. The purpose of this study was to compare the effects of CW and pulsed laser light on nerve regeneration in a hemisection model of SCI.

Materials and methods

This study was approved by the Institutional Animal Care and Use Committee, Uniformed Services University of Health Sciences. The study animals comprised 15 female Sprague-Dawley rats (200–250 g; Taconic Farms, Germantown, NY) which were randomized into the following three experimental groups: control, treatment with CW laser light, and treatment with pulsed laser light (PW). All rats underwent SCI by dorsal hemisection of the spinal cord at the T9 vertebral level as described previously [2, 3]. This dorsal hemisection model of SCI results in complete transection of the corticospinal tract, a major descending tract connecting the cerebral cortex with the motor neurons in the spinal cord that plays a dominant role in paralysis after SCI.

Within 15 minutes after SCI surgery, the CW and PW groups were transcutaneously irradiated with an 808-nm diode laser (provided by PhotoThera). Irradiation was delivered via a 3-mm diameter probe with direct contact to the skin above the lesion. Both treatment groups were treated at a power density of 0.5 W/cm² on the skin surface and an energy density of 1,500 J/cm². For the CW group, the output power was 35 mW. For the PW group, the

X. Wu · H. Moges · J. Anders (✉)
Department of Anatomy, Physiology and Genetics,
Uniformed Services University of the Health Sciences,
4301 Jones Bridge Road,
Bethesda, MD 20814, USA
e-mail: janders@usuhs.mil

L. DeTaboada
PhotoThera Inc,
Carlsbad, CA 92008, USA

parameters of the light were 175 mW peak power with 20% duty cycle, and a frequency of 100 Hz. Our previous study showed that irradiation with the same power density and energy did not induce significant heating at either the skin level ($1.832 \pm 0.068^\circ\text{C}$) or the spinal cord level ($0.350 \pm 0.018^\circ\text{C}$) [2]. Animals were irradiated for 50 min daily for 14 consecutive days. The total survival time for all rats was 3 weeks.

Ten days before the animals were killed, anterograde labeling was performed in all groups. A total of 12 μl 5% tetramethylrhodamine biotinylated dextran (mini-ruby; Molecular Probes, Carlsbad, CA) was injected into the primary motor cortex with six different coordinates (from bregma: -0.11 AP and ± 1.60 ML, -1.33 AP and ± 1.50 ML, -2.85 AP and ± 1.40 ML) [2]. This fluorescence dye labels the axons in the corticospinal tract located in the white matter [2, 3]. Axons were counted in 20- μm thick longitudinal sections of the spinal cord that extended from the lesion site to 20 mm caudal to the lesion using a TRITC filter (excitation 528–553 nm) and $\times 20$ magnification using a Nikon Labophot fluorescence microscope (Tokyo, Japan). Total axon number and average axonal length were calculated as described previously [2].

A footprint test was used as a functional assessment for this study. Briefly, animals walked along a ramp with their hind paws inked. Their footprint pattern was recorded on a strip of paper and measured for angle of rotation (angle formed by the intersection of the line through the prints of the third digit and the line through the central pad parallel to the walking direction). Only angle of rotation was measured in this study because our previous research showed that other factors, such as stride length and base of support, are not affected in this model of SCI [2, 3]. This test was performed prior to injury (as baseline) and on days 7, 14 and 21 after injury.

All statistical tests were performed using the GraphPad Prism program (GraphPad Software, San Diego, CA). One-way ANOVA was used to compare groups and Student's *t*-test was used for comparison between individual groups. Significant difference was defined as a *p* value less than 0.05.

Results

There were significantly higher total axon numbers in both the CW group and the PW group (504.0 ± 150.6 and 724.8 ± 78.9 , respectively) compared to the control group (111.3 ± 47.4 , $p < 0.05$ and $p < 0.01$, respectively; Fig. 1a). No significant difference was found between the CW group and the PW group. Distal to the lesion site, the CW group and the PW group had significantly greater average lengths of axonal regeneration (10.28 ± 0.81 and 13.54 ± 0.70 mm,

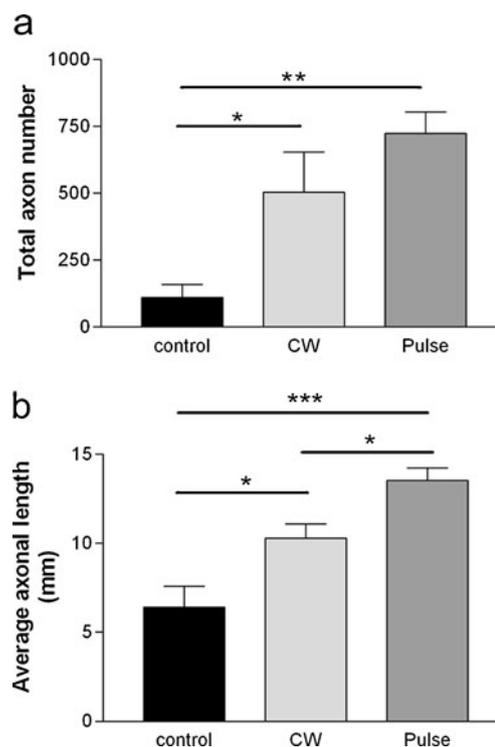


Fig. 1 Assessment of regrowing axons after SCI in control and light-treated groups: **a** total axon number; **b** average axonal length distal to the lesion site. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Error bars represent means \pm SEM

respectively) compared to the control group (6.42 ± 1.18 mm, $p < 0.05$ and $p < 0.001$, respectively; Fig. 1b). Also axonal length distal to the lesion site was significantly greater in the PW group than in the CW group ($p < 0.05$).

Behavioral assessment showed a significant difference in angle of rotation in both the CW group and the PW group compared to the control group ($p < 0.01$) when normalized to each group's baseline at 21 days after injury (Fig. 2). This reduction in angle of rotation indicated functional

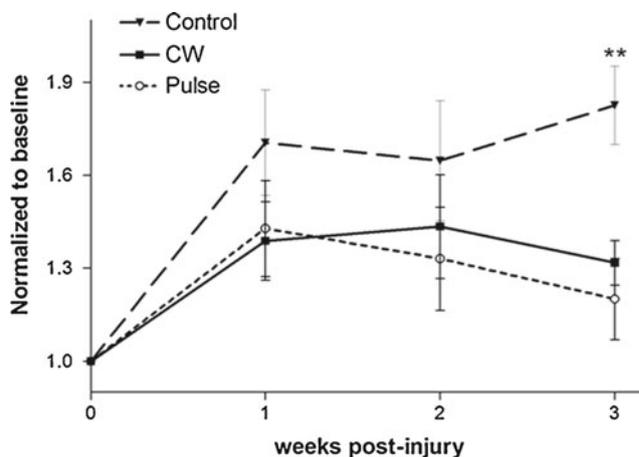


Fig. 2 Footprint test as functional analysis. ** $p < 0.01$. Error bars represent means \pm SEM

recovery after SCI. No significant difference was found between the CW group and the PW group.

Discussion

A number of recent reports on the comparison of the effects of pulsed versus CW light indicate that pulsed light may be more effective for LT applications than CW light. This potential advantage has stimulated interest and research resulting in a number of recent reports on the effects of pulsed light in LT.

CW light irradiation results in axonal regeneration in both the peripheral [15–17] and central injured nervous systems [2, 3]. As for LT using pulsed light for peripheral nerve injury, a few studies have shown either no effect or a worse outcome compared to no treatment [18, 19]. This study is the first to assess the efficacy of pulsed light in LT for SCI. The data showed that pulsed laser light significantly improved axonal regeneration in SCI compared to untreated controls and led to a significant increase in the length of regenerated axons distal to the lesion compared to the group treated with CW laser light ($p < 0.05$). Recent studies on neurological diseases have provided some insight into the possible mechanism involved in the beneficial effects of CW and pulsed light treatment. Transcranial laser treatment using CW light in rabbits following embolic stroke resulted in an increase in cortical ATP of 41% and pulsed light with 5 times and 35 times more energy delivery resulted in increases in cortical ATP of 157% and 221%, respectively [9]. Higher ATP content and improved mitochondrial function have also been found in a transgenic mouse model of Alzheimer's disease following transcranial irradiation using pulsed light [11]. Increased ATP content and enhanced mitochondrial function could be an acute effect which ultimately leads to normalization of the tissue function and neurological improvement.

To date, there is no established optimal range of parameters for pulsed light in LT. Simply following the effective parameters established for CW light irradiation does not ensure an efficacious treatment. Optimization of parameters for pulsed light needs to include frequency, duration time and duty cycle, besides the other parameters typically used in CW light irradiation, such as wavelength, output power, and power and energy density. In a rabbit embolic stroke model, laser treatment with pulsed laser light at two different frequencies (100 Hz and 1000 Hz) initiated 6 h after stroke significantly improved behavioral function. However, CW laser light with the same power density (7.5 mW/cm^2) and treatment time (2 min) did not result in a statistically significant improvement [6]. In a rat stroke model produced by permanent middle cerebral artery occlusion, CW light irradiation resulted in significant

functional improvement when applied 24 hours after stroke, while pulsed light irradiation at a frequency of 70 Hz resulted in a nonsignificant difference [7]. Although these two studies used different animal models of stroke, the difference in frequency of the pulsed light could be a main reason for the differences in the results. In this study, the parameters for the pulsed laser light were not optimized. The pulse parameters chosen were based on the same power density as used for CW laser light irradiation. The results of this study indicate a greater efficacy of pulsed laser light for repair of SCI. A follow-up study is planned to optimize the parameters of the pulsed light for clinical translation as a treatment for SCI.

For LT of a cutaneous or subcutaneous target, penetration depth is not a major issue. However, to treat diseased or injured tissues located deep inside the body, penetration depth is a critical factor. Depth of penetration depends upon the wavelength of the light used due to scattering and absorption of the light by tissue [20]. Therefore, if the same wavelength is used, CW and pulse modes of light will have the same penetration depth. However, the effective depth of penetration is determined by the depth where an adequate amount of energy is present to cause a photobiological reaction. Ilic et al. found that pulsed light could deliver 100 times the optimal power density without neurological damage at the cortical surface, while CW mode caused neurological deficits, including subdural necrosis with loss of neuronal tissue, at such a high power density [21]. The higher safe range of output power in pulsed light allows a greater effective penetration depth than CW mode. Hence, pulsed light can be used to treat not only a deeper target, such as nerves and muscles, but also the same target in larger organisms, which would allow the effective translation of LT from animal models to use in the clinic.

In conclusion, CW and pulsed laser light support axonal regeneration and functional recovery after SCI. Pulsed laser light has the potential to support axonal regrowth to spinal cord segments located farther from the lesion site. Therefore, the use of pulsed light is a promising non-invasive therapy for SCI.

Acknowledgments This study was supported by a Cooperative Research and Development Agreement between the Uniformed Services University of the Health Sciences and PhotoThera, Inc.

References

1. Rochkind S, Shahar A, Amon M, Nevo Z (2002) Transplantation of embryonal spinal cord nerve cells cultured on biodegradable microcarriers followed by low power laser irradiation for the treatment of traumatic paraplegia in rats. *Neurol Res* 24(4):355–360

2. Byrnes KR, Waynant RW, Ilev IK, Wu X, Barna L, Smith K, Heckert R, Gerst H, Anders JJ (2005) Light promotes regeneration and functional recovery and alters the immune response after spinal cord injury. *Lasers Surg Med* 36(3):171–185. doi:10.1002/lsm.20143
3. Wu X, Dmitriev AE, Cardoso MJ, Viers-Costello AG, Borke RC, Streeter J, Anders JJ (2009) 810 nm Wavelength light: an effective therapy for transected or contused rat spinal cord. *Lasers Surg Med* 41(1):36–41. doi:10.1002/lsm.20729
4. Oron A, Oron U, Streeter J, de Taboada L, Alexandrovich A, Trembovler V, Shohami E (2007) Low-level laser therapy applied transcranially to mice following traumatic brain injury significantly reduces long-term neurological deficits. *J Neurotrauma* 24(4):651–656. doi:10.1089/neu.2006.0198
5. Lapchak PA, Wei J, Zivin JA (2004) Transcranial infrared laser therapy improves clinical rating scores after embolic strokes in rabbits. *Stroke* 35(8):1985–1988. doi:10.1161/01.STR.0000131808.69640.b7
6. Lapchak PA, Salgado KF, Chao CH, Zivin JA (2007) Transcranial near-infrared light therapy improves motor function following embolic strokes in rabbits: an extended therapeutic window study using continuous and pulse frequency delivery modes. *Neuroscience* 148(4):907–914. doi:10.1016/j.neuroscience.2007.07.002
7. Oron A, Oron U, Chen J, Eilam A, Zhang C, Sadeh M, Lampl Y, Streeter J, DeTaboada L, Chopp M (2006) Low-level laser therapy applied transcranially to rats after induction of stroke significantly reduces long-term neurological deficits. *Stroke* 37(10):2620–2624. doi:10.1161/01.STR.0000242775.14642.b8
8. DeTaboada L, Ilic S, Leichliter-Martha S, Oron U, Oron A, Streeter J (2006) Transcranial application of low-energy laser irradiation improves neurological deficits in rats following acute stroke. *Lasers Surg Med* 38(1):70–73. doi:10.1002/lsm.20256
9. Lapchak PA, De Taboada L (2010) Transcranial near infrared laser treatment (NILT) increases cortical adenosine-5'-triphosphate (ATP) content following embolic strokes in rabbits. *Brain Res* 1306:100–105. doi:10.1016/j.brainres.2009.10.022
10. Moges H, Vasconcelos OM, Campbell WW, Borke RC, McCoy JA, Kaczmarczyk L, Feng J, Anders JJ (2009) Light therapy and supplementary riboflavin in the SOD1 transgenic mouse model of familial amyotrophic lateral sclerosis (FALS). *Lasers Surg Med* 41(1):52–59. doi:10.1002/lsm.20732
11. De Taboada L, Yu J, El-Amouri S, Gattoni-Celli S, Richieri S, McCarthy T, Streeter J, Kindy MS (2011) Transcranial laser therapy attenuates amyloid- β peptide neuropathology in amyloid- β protein precursor transgenic mice. *J Alzheimers Dis* 23:521–535. doi:10.3233/JAD-2010-100894
12. Lampl Y, Zivin JA, Fisher M, Lew R, Welin L, Dahlof B, Borenstein P, Andersson B, Perez J, Caparo C, Ilic S, Oron U (2007) Infrared laser therapy for ischemic stroke: a new treatment strategy: results of the NeuroThera Effectiveness and Safety Trial-1 (NEST1). *Stroke* 38(6):1843–1849. doi:10.1161/STROKEAHA.106.478230
13. Zivin JA, Albers GW, Bornstein N, Chippendale T, Dahlof B, Devlin T, Fisher M, Hacke W, Holt W, Ilic S, Kasner S, Lew R, Nash M, Perez J, Rymer M, Schellinger P, Schneider D, Schwab S, Veltkamp R, Walker M, Streeter J (2009) Effectiveness and safety of transcranial laser therapy for acute ischemic stroke. *Stroke* 40(4):1359–1364. doi:10.1161/STROKEAHA.109.547547
14. Hashmi JT, Huang YY, Sharma SK, Kurup DB, De Taboada L, Carroll JD, Hamblin MR (2010) Effect of pulsing in low-level light therapy. *Lasers Surg Med* 42(6):450–466. doi:10.1002/lsm.20950
15. Anders JJ, Borke RC, Woolery SK, Van de Merwe WP (1993) Low power laser irradiation alters the rate of regeneration of the rat facial nerve. *Lasers Surg Med* 13(1):72–82
16. Anders JJ, Geuna S, Rochkind S (2004) Phototherapy promotes regeneration and functional recovery of injured peripheral nerve. *Neurol Res* 26(2):233–239. doi:10.1179/016164104225013914
17. Gigo-Benato D, Geuna S, Rochkind S (2005) Phototherapy for enhancing peripheral nerve repair: a review of the literature. *Muscle Nerve* 31(6):694–701. doi:10.1002/mus.20305
18. Chen YS, Hsu SF, Chiu CW, Lin JG, Chen CT, Yao CH (2005) Effect of low-power pulsed laser on peripheral nerve regeneration in rats. *Microsurgery* 25(1):83–89. doi:10.1002/micr.20079
19. Bagis S, Comelekoglu U, Coskun B, Milcan A, Buyukakilli B, Sahin G, Ozisik S, Erdogan C (2003) No effect of Ga-As (904 nm) laser irradiation on the intact skin of the injured rat sciatic nerve. *Lasers Med Sci* 18(2):83–88. doi:10.1007/s10103-003-0258-6
20. Baxter DG (1994) *Therapeutic lasers: theory and practice*. Churchill Livingstone, London
21. Ilic S, Leichliter S, Streeter J, Oron A, DeTaboada L, Oron U (2006) Effects of power densities, continuous and pulse frequencies, and number of sessions of low-level laser therapy on intact rat brain. *Photomed Laser Surg* 24(4):458–466. doi:10.1089/pho.2006.24.458